

TruVivo[®] Application Brief: A Human-Relevant Model for CYP2C Induction and DDI Risk Assessment

The Research Challenge

Regulatory is placing more pressure on pharma companies to understand the translation of *in vitro* positive CYP3A4 induction results to other pathways co-regulated with CYP3A including other CYPs, UGTs and transporters. **Standard hepatocyte monoculture models fail** to capture the clinically observed induction potential of **CYP2C enzymes (CYP2C8, CYP2C9, CYP2C19)**.

Dynamic range is limited (< 2-fold mRNA increase for rifampicin) – forcing industry to infer CYP2C risk from CYP3A4 data. This has led to **underprediction of potential clinical DDI risk** and uncertainty in regulatory submissions.

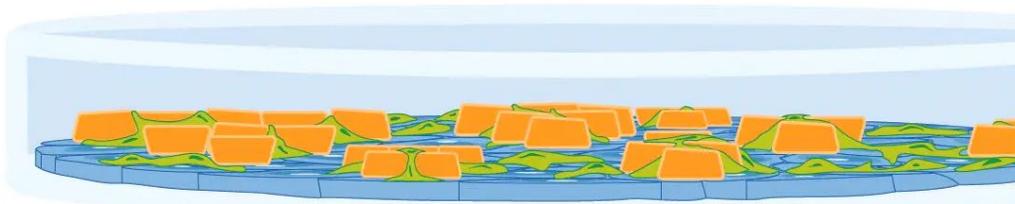
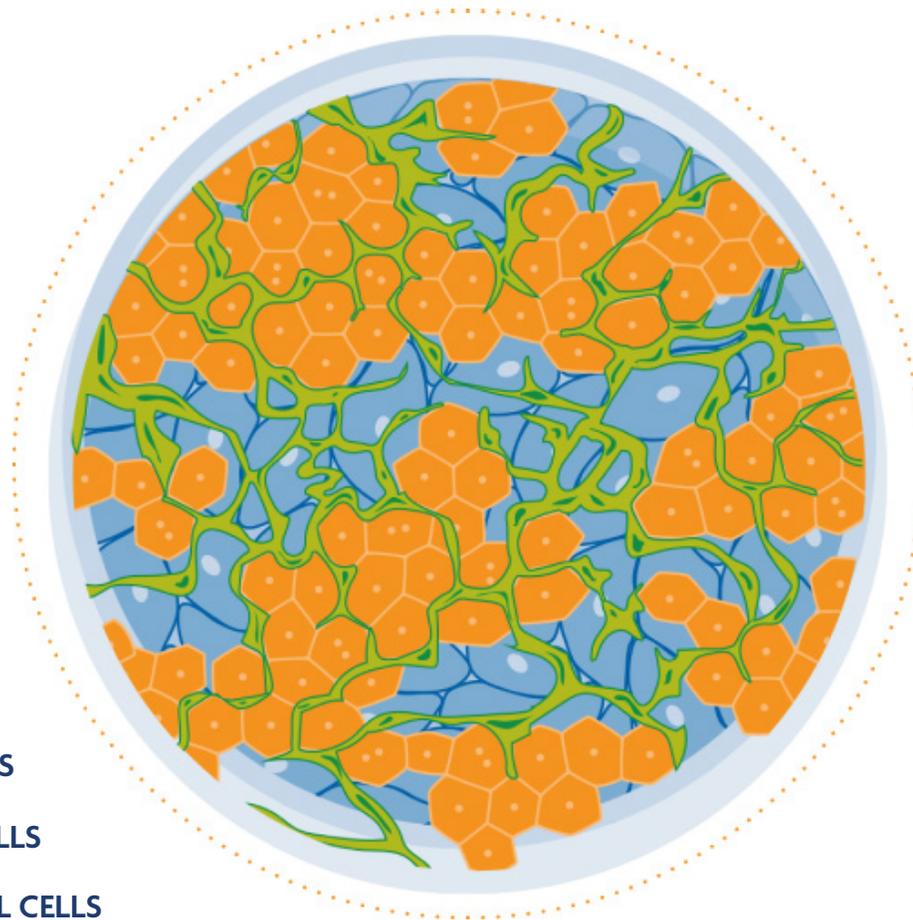
Researchers need a model that more accurately represents human CYP2C induction dynamics and allows IVIVE and PBPK modeling approaches to inform clinical outcomes.

Figure legend: Schematic representation of TruVivo showing top and side view of the self-assembled, multicellular hepatocyte colonies integrated among a mixed feeder cell layer.

The TruVivo[®] Solution

An advanced human hepatic triculture model of all-human primary cells — hepatocytes supported by stromal and endothelial feeder cells — delivering sustained function, reproducibility, and significantly improved nuclear receptor signaling for generating accurate, reproducible CYP induction data.

-  HEPATOCYTES
-  STROMAL CELLS
-  ENDOTHELIAL CELLS



Key Findings and Validation

Finding	Evidence
Robust and reproducible CYP2C induction	<ul style="list-style-type: none">• Rifampicin, apalutamide, efavirenz, and carbamazepine produced concentration-dependent increases in CYP2C8, CYP2C9, and CYP2C19 mRNA and activity across three donors².• E_{max} up to 8–20× (CYP2C8/19) and 2–4× (CYP2C9) — exceeding the <2-fold dynamic range of conventional hepatocyte systems².
Improved CYP2C19 expression vs. 2D culture	<ul style="list-style-type: none">• TruVivo maintained CYP2C19 gene expression 4–6× higher than 2D sandwich culture hepatocytes, enabling quantifiable induction responses not measurable in standard systems².
Quantitative translation to clinical outcomes*	<ul style="list-style-type: none">• TruVivo-derived induction parameters achieved AUCR predictions within 2-fold of clinical data for >80% of CYP2C substrates².• For CYP2C9 and CYP2C8, 85% and 64% of induction predictions captured $\geq 80\%$ of observed clinical effects¹.• Supported accurate IVIVE for CYP3A4, CYP2C19, UGT1A4, and P-gp induction pathways¹.
PBPK modeling confirms predictive performance	<ul style="list-style-type: none">• Integration of TruVivo data into Simcyp PBPK models produced excellent alignment with clinical pharmacokinetic profiles².• Outperformed mechanistic static models (MSM) by reducing over- and under-prediction of DDI magnitude².• Enabled full concentration–time profile prediction for victim drugs such as omeprazole, tolbutamide, and glyburide².
Cross-laboratory reproducibility	<ul style="list-style-type: none">• Independent studies demonstrated <2-fold variability in CYP2C induction response across laboratories using TruVivo¹ — compared to >10-fold variability typical of 2D hepatocyte cultures.
Multi-pathway and complex DDI modeling	<ul style="list-style-type: none">• TruVivo accurately predicted net induction and inhibition outcomes for co-inducer/inhibitor compounds (e.g., ritonavir)¹.• Maintained PXR/CAR signaling fidelity, allowing concurrent evaluation of CYP2C, CYP3A4, UGT, and transporter induction¹.
Consistent donor performance	<ul style="list-style-type: none">• CYP2C induction EC_{50} and E_{max} values were highly consistent across donors, minimizing inter-donor variability².• Stable phenotype and nuclear receptor responsiveness sustained throughout extended culture².

*CYP2C19, CYP3A4, UGT1A4, and P-gp all showed quantitative IVIVE correlation, typically within 2-fold of observed AUCRs once GMFE correction was applied for CYP2C19 and CYP3A4.

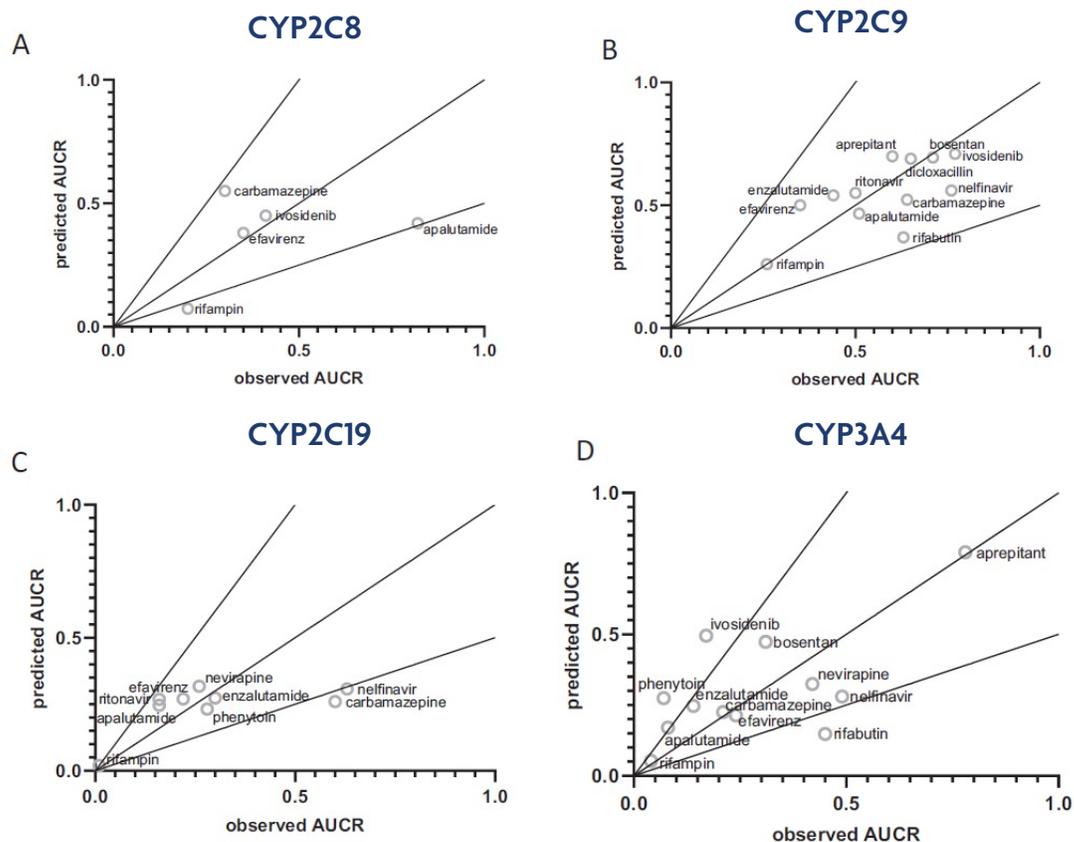


Figure legend: Predictive accuracy of *in vitro*-*in vivo* extrapolation (IVIVE) for CYP2C8, CYP2C9, CYP2C19, and CYP3A4

Model predictions using averaged donor (n=2) data closely align with clinical outcomes, demonstrating strong translational performance. The central line shows perfect prediction, with upper and lower lines marking a two-fold range. GMFE corrections were applied for CYP2C19 and CYP3A4 to refine accuracy. Figure reproduced from portions of Figure 6 of Ramsden et al. with permission under an open access license.¹

Why Researchers Choose TruVivo

Researchers Need

TruVivo Advantage

Predictive Accuracy

Clinically aligned CYP2C induction responses — AUCR predictions within 2-fold of observed clinical outcomes for >80% of test compounds, including CYP2C8, CYP2C9, CYP2C19, CYP3A4, UGT1A4, and P-gp^{1,2}.

Nuclear Receptor Biology

Authentic human cellular composition and architecture sustain **endogenous PXR and CAR activation**, driving physiologic regulation of CYP2C isoforms and accurate induction magnitude^{1,2}.

Long-term Stability

Maintains hepatocyte phenotype and enzyme function for **multi-day exposures and repeated dosing**, enabling concentration-response studies and chronic DDI modeling².

Multi-enzyme and Transporter Insight

Parallel evaluation of **CYP2C, CYP3A, UGT, and P-gp** induction within the same culture — supporting integrated mechanistic modeling of hepatic metabolism and clearance¹.

Cross-lab Reproducibility

Demonstrated **<2-fold variability** in induction magnitude across independent laboratories, compared with >10-fold variability typical of 2D hepatocyte systems¹.

Complex DDI Modeling Capability

Successfully predicted **net induction and inhibition outcomes** for dual-acting compounds such as **ritonavir**, enabling realistic assessment of competing metabolic pathways¹.

Industry Validation

Independently evaluated and published by leading pharma collaborators (AstraZeneca, IQ Induction Working Group) as a reliable tool for **quantitative IVIVE of CYP2C-mediated DDI risk**^{1,2}.

Research Applications

- Induction risk assessment for **CYP2C enzymes** (CYP2C8/9/19)
- **Clinical DDI prediction** and quantitative IVIVE modeling
- Mechanistic studies of **nuclear receptor activation pathways**
- Early de-risking for **co-inducer/inhibitor compounds**

References

1. **Ramsden D.**, Fullenwider C.L., Santos C., LeCluyse E.L. (2025) Quantitative Clinical Risk Assessment of CYP2C, UGT, and P-gp Induction Using the TruVivo Human Hepatocyte Triculture Platform. Drug Metab Dispos 2025.
2. **Slavsky M.**, Karve A.S., Hariparsad N. Physiologically Based Pharmacokinetic Modeling to Assess Perpetrator and Victim CYP2C Induction Risk. Pharmaceutics 2025, 17(1085).



Learn how TruVivo can transform your DDI and induction studies.

LifeSciences@lifenethealth.org | LNHLifeSciences.org/TruVivo

